# In vitro Indicators of Moringa oleifera Nutritional Value for Ruminants in the Dry Season

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#### ABSTRACT

*In vitro* indicators of *Moringa oleifera* nutritional value for ruminants during the dry season in Camagüey municipality, Cuba, were determined. This species is abundant in Santayana, Imán, and La Belén areas. Despite significant differences in raw protein (23,6 % and 23,0 %) and neuter-detergent fibre (17,0 % and 25,9 %), none were found for dry matter (20,0 % and 23,2 5) and ashes (7,8 5 and 10,0 %). raw protein and dry matter highest values were registered in La Belén, while the lowest ones were detected in Santayana. Neither saponins nor alkaloids were present in the three areas; however, tannins and condensed tannins had a higher prevalence in Santayana. *In vitro* gas production out of bovine feces shoed significant differences at 12; 24, and 96 hrs of incubation. *M. oleifera* nutritional value is high in these areas.

Key Words: chemical composition, in vitro gas production, ruminants

#### **INTRODUCTION**

*Moringa oleifera* has stood out within the nonlegume trees as a promising plant for animal nutrition. Its leaves are avidly consumed by all kinds of animals: ruminants, camels, pigs, carps, tilapias and other herbivore fishes. Hence, it is considered one of the "complete" forages (very rich in protein, vitamins and minerals and high palatability) (Moroto *et al.*, 2000).

The *in vitro* indicators of nutritive values of *Moringa oleifera* foliage from three different areas were determined for ruminants in the municipality of Camagüey, during the dry season.

#### MATERIALS AND METHODS

*Moringa oleifera* leaves, stems, and petioles were chosen at random in the morning (7:00-9:00 am) with a diameter lower than five millimeters, from five plants used in hedges in three different locations in the municipality of Camagüey, Cuba: Santayana (21° 20' 15'' N and 77° 50' 55'' O), on Dark red fersialitic soil; Imán (21° 23' 25'' N and 77° 57' 30'' O) and La Belén (21° 22' 05'' N and 77° 55' 40'' O), both on brown soil without carbonate. The *M. oleifera* samples were collected in Santayana and Imán from 120 dayold plant reshoots; and 60 day-old reshoots from plants in La Belen. Sampling was carried out in February 2012.

The plants were quickly taken to the lab, sliced and homogenized; then they were dried at 66°C to achieve constant weight, for about 48 h in an electric oven with forced air circulation, then the samples were crushed through a 1 mm sieve.

The presence of antinutritive substances and analyses of dry matter (MS), ash, and gross protein (PB) were made by triplicate, according to AOAC (1995); whereas neutral detergent fiber determination (FDN) was done using the Van Soest and Robertson (1985) method.

The general procedure used for *in vitro* gas production was based on the principles set up by Menke *et al.* (1979) with the use of 100 ml glass syringes (1 ml appreciation). Bovine feces were used as inoculum (Martínez, 2008); the feces were gathered early in the morning before three hours after deposition; then they were mixed with the buffered mineral medium. All the procedure was performed with systematic injection of  $CO_2$ .

The initial volume (Vo) of each syringe was recorded and then placed in Luke-warm water bath, at 39 °C. Readings were made at 3; 6; 12; 24; 48; 72 and 96 h. Three syringes with inoculums + buffer were prepared to be used as target; other three syringes were used as guinea (*Pannicum maximum*) sample patterns. All samples were incubated by triplicate and the volume of gas was expressed as gas milliliter every 200 mg of dry samples.

The parameters for the production of *in vitro* gas were determined using the Ørskov and McDonald (1979) equation:

p = A + B \* [1 - EXP (-C \* t)]

Where:

t: time (h).

A: volume at 0 h.

B: gas volume (ml).

A + B: gas production potential.

C: specific speed of gas production in the exponential phase  $(h^{-1})$ .

Data normalcy was verified and descriptive statistics was performed. The composition of the chemical value and gas production in every hour of incubation were compared among the sample origins, by simple variance analysis. The differences between the means were determined by the Tukey test. Al the analysis was performed with SPSS software version 17.0.

### **RESULTS AND DISCUSSION**

Table 1 shows the chemical composition indicators from the three areas where *M. oleifera* was collected. Dry matter and ash were observed to have no significant differences among them; contrary to PB and la FDN. Moreover, the samples from La Belen had the highest values of PB and MS; the lowest were found in Santayana. However, the samples from Santayana had the highest percents of ash and FDN compared to La Belen.

Chemical composition variations in pastures and forages are well known to be associated with several different factors related to soil, the weather and plant handling. In this case, the differences among the origins may be influenced by a combination of factors where soil humidity and fertility are significant. Handling factors are mainly related to the time and kind of trimming. The samples from La Belen were observed to reshoot in fewer days, which has been associated to higher concentrations of PB and lower of fiber (Pedraza, 2000); however, fiber composition is known to determine digestibility (Goering and Van Soest, 1970), which must be considered in future studies.

Table 2 shows the presence of nutritional factors in foliage. Tannins and condensed tannins were found in all of it, more in Santayana. No saponin or alkaloids were found in any.

The antinutritional factors are important for plants, as the only defense mechanism against natural predators (Reed, 1995). In addition, this may become a disadvantage, when the levels are relatively harmful, some of these factors may lower protein digestibility (Huisman, 1991; Makkar and Becker, 1996) and affect ruminal fermentation. (Rodríguez *et al.*, 2011); others, like tannins in low concentrations, may increase protein use in rumen, or contribute with the reduction of intestinal parasites (Min and Hart, 2003).

The amount of gas produced depends on the quality, digestibility and energy value of the feed assessed (Menke *et al.*, 1979; Ammar *et al.*, 2005; Posada and Noguera, 2005).

Significant differences (P < 0.05) among the origins were observed only at 12, 24 and 96 h of incubation. The highest values of cumulated gas were observed in the foliage from La Belen; whereas the collections from Imán and Santayana do not differ between them for the total cumulated production, at 96 h.

Figure 1 shows the *in vitro* gas production patterns, according to the equation of Ørskov and McDonald (1979). La Belen collections had the highest production of gas in comparison with Imán and Santayana. The gas production speeds were similar.

## **CONCLUSIONS**

El valor nutritivo del follaje de las tres procedencias de *M. oleifera* para rumiantes es alto.

#### **R**EFERENCES

- AMMAR, H.; LÓPEZ, S. y GONZÁLEZ, J. S. (2005). Assessment of the Digestibility of Some Mediterranean Shrubs by *in vitro* Techniques. *Animal Feed Science and Technology*, 5 (3), 323-331.
- ASSOCIATION of OFFICIAL ANALYTICAL CHEMIST (1995). *Official Methods of Analysis* (16<sup>th</sup> Edition). Washington, DC.: AOAC International.
- GOERING, H. K. y VAN SOEST, P. J. (1970). Forage Fiber Analysis. En Agriculture Handbook. Washington, D.C., USA: Agricultural Research Service-USDA.
- HUISMAN, J. (1991). Negative Effects of Antinutritional Factors in Animal Feeds on the Excretion of Nitrogen in the Environment and Possibilities of Reduction. Digestive Physiological Effects. *Tijdschrift Voor Diergeneeskunde 116* (8), 391-395.
- Makkar, H. y Becker, K. (1996). Nutrients and Antiquality Factors in Different Morphological Parts of the *Moringa oleifera* Tree. *Journal of Agricultural Science*, 67 (45). Cambridge.
- MARTÍNEZ, S. J. (2008). Heces vacunas depuestas como inoculo en la técnica de producción de gases para la valoración nutritiva in vitro de forrajes.

Trabajo de doctorado en Ciencias Veterinarias, Universidad de Camagüey, Cuba.

- MENKE, K. H.; RAAB, L.; SALEWSKI, A.; STEINGASS, H.; FRITZ, D. y SCHNEIDER, W. (1979). The Estimation of the Digestibility and Metabolizable Energy Content of Ruminant Feedingstuffs from the Gas Production when they are Incubated with Rumen Liquor *in vitro*. J. Agric. Sci., 93 (21), 217-222.
- MIN, B. R. y HART, S. P. (2003). Tannins for Suppression of Internal Parasites. J. Anim. Sci., 81, 102-109.
- MOROTO, L. O.; CRUZ, E., FRANCAISE, E.; DRIESCHE, V.; BECKMANS, S.; MANSO, M. J.; LAZO, L.; RÍOS, C. y MACHADO, J. M. (2000). Moringa oleifera *Lam* (Pterigosperma): *Consideraciones sobre la presencia de lectinas*. IV Taller Internacional Silvopastoril Los árboles y arbustos en la ganadería tropical, Estación Experimental de Pastos y Forrajes *Indio Hatuey*, Matanzas, Cuba.
- ØRSKOV, E. R. y MCDONALD, I. (1979). The Estimation of Protein Degradability in the Rumen from Incubation Measurements Weighted According to Rate of Passage. Cambridge. *Journal of Agricultural Science*, 92, 499-503.

- PEDRAZA, R. M. (2000). Valoración nutritiva del follaje de Gliricidia sepium (Jacq) Kunth ex Walp y su efecto en el ambiente ruminal. Trabajo de doctorado en Ciencias Veterinarias, Instituto de Ciencia Animal-Universidad Agraria de La Habana, Cuba.
- POSADA, S. L. y NOGUERA, R. R. (2005). Técnica in vitro de producción de gases: Una herramienta para la evaluación de alimentos para rumiantes. Livestock Research for Rural Development, 17 (4). Extraído el 25 de mayo de 2012, desde http://www.cipav.org.co/lrrd/lrrd17/4/posa17036.
- REED, J. D. (1995). Nutritional Toxicology of Tannins and Related Polyphenols in Forages Legumes. *Journal of Animal Science*, 73 (5), 56-60.
- RODRÍGUEZ, R.; BRITOS, A.; RODRÍGUEZ-ROMERO, N. y FONDEVILA, M. (2011). Effect of Plant Extracts from Several Tanniferous Browse Legumes on *in* vitro Microbial Fermentation of the Tropical Grass Pennisetum purpureum. Anim. Feed Sci. Technol., 168, 188-195.
- VAN SOEST P. J. y ROBERTSON, J. B. (1985). Analysis of Forages and Fibrous Foods. A Laboratory Manual for Animal Science 613. Report of Research of the Cornell University Agricultural Experiment Station, 35 (3), 2-3.

Received: 10-6-13 Accepted: 10-7-13

Factors antinutritio-	Origins					
nals	Santayana	Imán	La Belén			
Tannnins	+	+	+			
Condensed Tannnins	++	+	+			
Saponins	-	-	_			
Alkaloids	_	-	_			
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Table 1. Presence of antinutritive factors in the three collections of *M. oleifera* 

(+) Presence

(-) Absence

Table 2. Influence of <i>M. oleifera</i> origin on <i>in-vitro</i>	gas production at different incubation times
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Times/incubation	Origins							
	Santayana	Imán	La Belén	ES	Sig.			
3	0.9	1.1	1.2	0.29	NS			
6	1.7	3.1	3.1	0.37	NS			
12	3.7 <sup>ac</sup>	5.8 <sup>b</sup>	6.9 <sup>a</sup>	0.57	*			
24	12.2 <sup>c</sup>	13.5 <sup>b</sup>	16.2 <sup>a</sup>	0.72	*			
48	17.8	19.9	21.8	0.87	NS			
72	23.0	23.6	27.4	0.94	NS			
96	27.5 <sup>b</sup>	27.6 <sup>b</sup>	32.2 <sup>a</sup>	0.93	*			

NS: no significant difference

\* P < 0.05

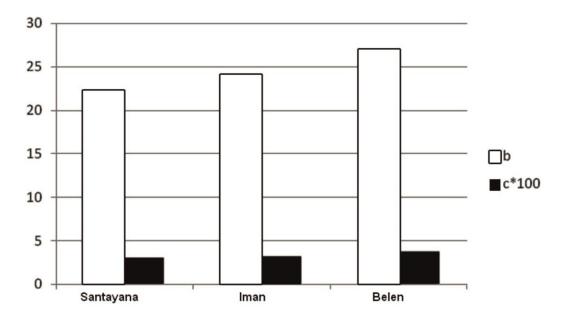


Fig. 1. In vitro gas production parameters (ml) of the three M. oleifera origins